Assessment of the conditions of the thermoplastic extrusion process in the bioactive and mechanical properties of flexible films based on starch and Brazilian pepper

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Abstract
The objective of this work was to produce, through the thermoplastic extrusion process followed by blowing, manioc starch-based flexible films added with Brazilian pepper oil as an antioxidant and plasticizer agent, and verify if the bioactive compounds contained in the fresh pepper oil are present after the drying step of the thermoplastic extrusion. After analysis by gas chromatography-mass spectrometry volatiles compounds were identified in the films. Pepper oil also influenced the mechanical properties of the films. These results suggest that the temperatures used in the process, kept some of the existing compounds in the Brazilian pepper essential oil adhered to the packages.

Keywords: Thermoplastic extrusion, temperature, bioactive compounds.
1. Introduction

Plastic films of petrochemical origin are currently used on a large scale in food packaging, and their use is contingent upon their mechanical and barrier properties. Although chemically stable, they are not biodegradable and cause environmental impact. One of the alternatives to this problem would be the development of edible and/or biodegradable films using raw materials from renewable sources.

The bioplastics industry is still in beginning, looking to identify and exploit market niches not only for the biodegradability, recycling and/or replacement of plastics of petrochemical origin by plastics from renewable sources, also concerned about the carbon cycle and sustainability. Bioplastics are defined as materials which, containing variable percentage biopolymers, can be molded by heat and pressure. They are potential alternatives to conventional thermoplastic polymers of petrochemical origin, such as polyolefins and polyesters \[1\].

Starch is a polysaccharide of plant origin, widely found in the international market, capable of forming films both by the casting technique and by the thermoplastic extrusion process. Films made with starch can also introduce additives into the packaging, such as natural antioxidant and antimicrobial agents.

The Brazilian pepper (Schinus terebinthifolius Raddi) is originally from Peru and is widely distributed throughout the Americas, including the Northeast, Southeast, South and Midwest of Brazil \[2\]. The fruit has healing properties attributed to the different volatile compounds that are distributed in its various organs, such as bark, leaves, flowers, fruits and seeds \[3\]. It is used in the food industries, as well as its essential oil in the development of pharmaceuticals and cosmetics \[4\]. The species has increasing pharmacological use, and is considered by popular medicine as an anti-inflammatory, antimicrobial and strong antioxidant agent \[4,5,6\].

The process of extrusion allows the thermoplastification of a solid material by the application of heat and mechanical work, being the main process to obtain the bioplastics. It is a highly versatile process in which the extruder can behave as a heat exchanger due to the thermal changes occurring between the material to be extruded and the equipment \[7\].

In this context, the objective of this work was to produce, through the thermoplastic extrusion process followed by blowing, manioc starch-based flexible films added with Brazilian pepper oil as an antioxidant and plasticizer agent, and verify if the bioactive compounds contained in the fresh pepper oil are present after the drying step of the thermoplastic extrusion process to obtain the biodegradable film. In parallel, we analyzed

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the mechanical properties of the films to verify the effect of the pepper oil concentration on tensile strength, elongation and elasticity of the films.

2. Materials and Methods

Materials: manioc starch, glycerol and brazilian pepper essential oil.

The films were produced by thermoplastic extrusion following buy blowing in single screw extruder. The range of temperature was 90 to 130°C and the velocity 35 rpm, the same conditions proposed by Fakhouri [8]. Three formulations were produced: i) manioc starch with glycerol (25%), ii) manioc starch, glycerol (25%) and brazilian pepper essential oil (2%) and iii) manioc starch, glycerol (25%) and brazilian pepper essential oil (3%).

Chemical characterization by Gas Chromatography: The analyzes were performed using a gas chromatograph equipped with a mass spectrometer detector (GCMS-QP2010 Ultra, Shimadzu, Kyoto, Japan). employing a fused silica DB-5 capillary column (I & W, 5% de phenyl-dimethylpolysiloxane) with 30 m in length x 0.25 mm i.d., 0.25 µm film thickness, under the following conditions: carrier gas helium (99.999% and flow rate 1.0 mL min⁻¹); 1 µL injection volume, split ratio (1:20), with initial oven temperature of 50°C and heating from 50°C to 280°C at 3°C min⁻¹. The injector temperature, transfer line and detector temperatures were 250°C. The MS scan parameters included electron impact ionization voltage at 70 eV, a mass range of 45 to 600 m/z and a scan interval of 0.3 s. Temperature-programmed retention indices were calculated using a mixture of alkanes (C8-C30) as external references and compared with Adams[9]. The identifications were completed by comparing the mass spectra obtained in the NIST21 and WILEY229 databases and literature data [8].

Total flavonoids quantification by Spectrophotometry: The flavonoids content was determined in the acetonic extracts of extruded S. terebinthifolius using the colorimetric method involving the reaction with aluminum chloride by Chang [10] with adaptations for extrudates. Extracts were prepared with 100 g of sample added in 500 mL of acetone (50% w/v), and they were kept under constant agitation (150 rpm) for five hours. The sample was filtered, and the filtrate was considered the flavonoid extract for analysis.

The extract was reacted with aluminum chloride and the readings were performed in a spectrophotometer (Biochrom – Libra S60) adjusted at 415 nm. Quercetin solutions at nine concentrations (0.01 to 0.2 µg.µL⁻¹) were reacted with sodium aluminum chloride in order to construct a standard curve. The results were expressed as milligrams of quercetin equivalent (QE mg.100g⁻¹ sample) using the quercetin standard curve.
Elastic modulus, tensile strength and elongation at rupture will be determined using a texturometer (Universal testing machine Galdabini SUN 2500, TA-HDi Texture Analyser (Stable Microsystem, Surrey, England), the conditions were perfomed by ASTM methods\cite{11}.

3. Results and Discussion

After analysis by gas chromatography-mass spectrometry, fourteen volatile compounds were found in the essential oil of S. terebinthifolius (Table 1). Among these compounds, we highlight the iso-sylvestrene (34.12%), α-thujene (17.13%), myrcene (8.25%), α-phellandrene (6.68%), β-longipinene (5.90%) and sylvestrene (5.87%). After the thermoplastic extrusion process at temperatures ranging from 90 to 130°C, 5 volatile compounds were identified, knowingly: α-pinene, sabinene, β-pinene, limonene and α-copaene.

<table>
<thead>
<tr>
<th>Compounds</th>
<th>RI\textsuperscript{a}</th>
<th>RI\textsuperscript{b}</th>
<th>Essential oil (%)</th>
<th>Extruded (2%)</th>
<th>Extruded (3%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>α-thujene</td>
<td>923</td>
<td>924</td>
<td>17.13</td>
<td>-</td>
<td>-</td>
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<tr>
<td>α-pinene</td>
<td>939</td>
<td>939</td>
<td>1.03</td>
<td>17.43</td>
<td>18.68</td>
</tr>
<tr>
<td>α-fenchene</td>
<td>953</td>
<td>953</td>
<td>3.83</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Sabinene</td>
<td>975</td>
<td>976</td>
<td>-</td>
<td>9.54</td>
<td>10.13</td>
</tr>
<tr>
<td>β-pinene</td>
<td>980</td>
<td>980</td>
<td>1.19</td>
<td>8.79</td>
<td>8.67</td>
</tr>
<tr>
<td>Myrcene</td>
<td>988</td>
<td>988</td>
<td>8.25</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>α-phellandrene</td>
<td>1002</td>
<td>1002</td>
<td>6.68</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Iso-sylvestrene</td>
<td>1007</td>
<td>1007</td>
<td>34.12</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Sylvestrene</td>
<td>1025</td>
<td>1025</td>
<td>5.87</td>
<td>-</td>
<td>-</td>
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<tr>
<td>Limonene</td>
<td>1030</td>
<td>1029</td>
<td>-</td>
<td>15.24</td>
<td>15.41</td>
</tr>
<tr>
<td>Fenchone</td>
<td>1083</td>
<td>1083</td>
<td>3.69</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>α-copaene</td>
<td>1377</td>
<td>1377</td>
<td>-</td>
<td>9.91</td>
<td>10.12</td>
</tr>
<tr>
<td>β-longipinene</td>
<td>1400</td>
<td>1400</td>
<td>5.90</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Aromadendrene</td>
<td>1465</td>
<td>1465</td>
<td>4.63</td>
<td>-</td>
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</tr>
<tr>
<td>Biciclogermacrene</td>
<td>1517</td>
<td>1517</td>
<td>3.21</td>
<td>-</td>
<td>-</td>
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<tr>
<td>δ-cadinene</td>
<td>1519</td>
<td>1519</td>
<td>2.29</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Sphatulenol</td>
<td>1619</td>
<td>1619</td>
<td>1.34</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

Total flavonoids (mg.100g QE) - - - 60.77 63.49

\textsuperscript{a}Retention index calculated; \textsuperscript{b}Retention index literature. The values of gas chromatography are expressed in % and the total flavonoids are expressed in mg.100g of quercetin equivalent.
Higher concentrations of pepper oil influenced the mechanical properties of the films, causing a significant increase in the elongation of the films and decrease of the tensile strength and elasticity (Figure 1).

**Figure 1.** (a) Young modulus (MPa), (b) Elogation at break (%) and (c) Tensile Strenght (MPa) of flexible films from starch and Brazilian pepper.
4. Conclusions

It was possible to obtain flexible films by manioc starch, glycerol and brazilian pepper essential oil by thermoplastic extrusion following buy blowing. After the thermoplastic extrusion process, five volatile compounds were identified, knowingly: α-pinene, sabinene, β-pinene, limonene and α-copaene in the flexible films. The brazilian pepper essential oil also influenced the mechanical properties of the films.

These results suggest that the temperatures used in the thermoplastic extrusion process, although high, kept some of the existing compounds in the Brazilian pepper essential oil adhered to the packages.

5. Acknowledgment

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6. References


